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Nano fertilizer is boon for Soil Health

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Article History

ABSTRACT

Received: 03/04/2021
Accepted: 06/05/2021Nanoparticles are being increasingly used in the agriculture sector as
opposed to conventional fertilizers. As it can be observed from the
results, the nanofertilizers used are quite beneficial for plant growth.
Biosynthesis of nanoparticles can be the cheaper method of
production since the technology for large scale growth of bacteria
and simple eukaryotes like yeasts is already well stabli shed.*Corresponding E-mail:
shobhawaghmode@gmail.comThough research on nanofertilizers offer great benefits to crop
production and at the same time do not pollute the environment.

1. INTRODUCTION -

Nanotechnology, the concept is of manipulating material on the nanoscale causing significant changes in its properties. This technology has been used in various sectors such as automobile, electronics, IT, pharmaceutical, medical and cosmetic sectors(WHO meeting) In the past two decades advent of nanotechnology is being seen in the agriculture sector, especially in terms of precision agriculture(Duhan et al.,2017) . Precision agriculture basically implies the use of technology for making precise decisions which help in improvement of crop quality and yield. In this aspect, nanofertilizers and nanopesticides are being used. This review will concentrate on application of nanofertilizers in modern agriculture.

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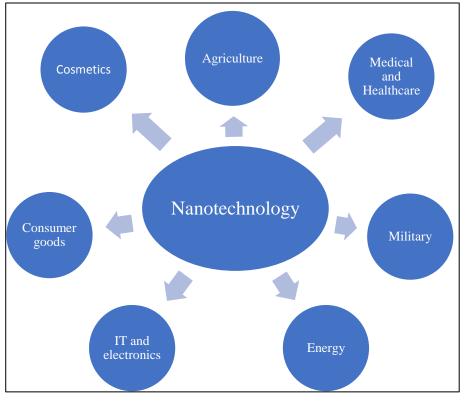


Figure 1: Uses of nanotechnology

Nanoparticles are materials less than 100nm in size. Owing to their small size, they have a high surface area to volume ratio. This ratio indicates that a large amount of nutrients can be attached to a relatively small amount of nanoparticles. This exact concept is used in manufacturing of nanofertilizers.

Conventional fertilizers like chemical fertilizers have an adverse effect on the environment as well as low uptake of nutrients. Environmental pollution includes leaching, de-nitrification and volatilization of chemical fertilizers. Leaching may lead to eutrophication while nitrogen volatilization causes release of nitrous oxide in the air. This being a greenhouse gas, contributes to global warming("FAO/WHO expert meeting on the application of nanotechnologies in the food and agriculture sectors," n.d.)

Inversely nanofertilizers have a high efficiency of nutrient uptake and almost no effect on the environment as indicated by various studies.In nanofertilizers, nanoparticles are used as carriers for micronutrients like zinc, copper, manganese, iron and macronutrients like nitrogen, phosphorous, potassium, sulphur and calcium(Dimkpa and Bindraban, 2018).

Uptake of these nanoparticles varies among different plant species, but also depends on the chemical composition, function and size of the nanoparticles. A review by Rico *et al*(2011). shows the different pathways for uptake for different nanoparticles. This is shown in a concise manner in fig.(1). As

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can be observed from the figure, the data is about the uptake of nanoparticles by plants. still insufficient to form a proper conclusion

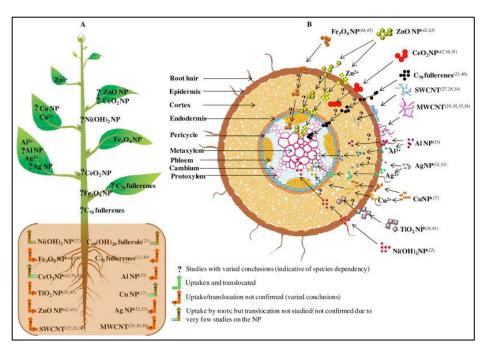


Figure 2: Uptake, translocation and biotransformation of various nanoparticles in a plant (Rico *et al.*,2011)

1.1 Toxicity -

A number of studies have been published regarding cytotoxicity of nanoparticles to plants as well as other ecological systems. As with toxicity studies, the result differs according to the conditions. It depends on the nanoparticle used, its quantity as well as the plant/s under study. Most of these studies show that nanoparticles used in proper amount and applied properly are not toxic to the plants and environment. This is seen from the fact that unlike conventional fertilizers, nanofertilizers do not volatilize and are almost completely absorbed by the plants various pathways as discussed above. However there are a few studies like the one by Wang et al.(2016) which shows that ZnO nanoparticles have a totally negative effect on *Arabidopsis* plants. Hence it is prudent to check toxicity of nanofertilizers before its large scale application.

2. MATERIALS AND METHODS -

Some of the most commonly used methods for synthesis and then characterization of nanoparticles used as nanofertilizers are stated in the following text.

2.1 Synthesis of Nanoparticles :

a)Chemical methods of synthesis -

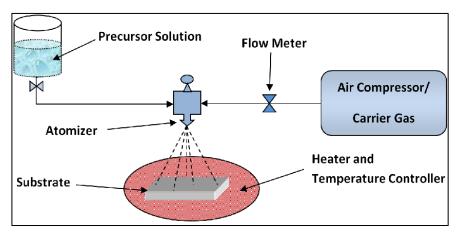
Spray pyrolysis : This is a simple method of nanoparticle synthesis which is cost effective and easy to perform*. In this technique, precursor solutions containing the desired nanoparticles are sprayed in the form of

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droplets on the hot surface of a furnace. This causes the solvent in the precursor solution to evaporate and leaves the nanoparticles in

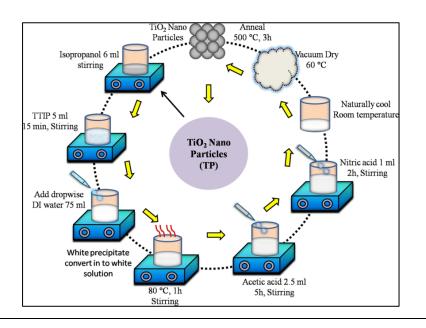
the vapour phase. A schematic representation of this process by Trakhtenberg *et al.* is shown in fig.(3)





Sol - gel method :

This technique is a wet chemical process so its starting material is a solution or "sol". Through this process, the solution get gradually polymerized to form a gel like structure or colloid. Pawar *et al.*(2019) describes the use of sol-gel method for formation of titanium oxide nanoparticles. Here, they have used vacuum dry as final step for densification and formation of the nanoparticles. Advantage of this process is that it does not require expensive equipment and is relatively simple to perform.



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Figure 4: Sol-gel method (Pawar *et al.*)

Hydrothermal method :

Hydrothermal method is commonly used to synthesize nanoparticles and is solution reaction-based method. This technique utilizes control of thermodynamic variables like temperature and pressure to form crystalline structures from solutions. The morphology and composition of the product nanoparticles depends on the settings of these variables along with the vapour pressure of the nanoparticles to be synthesized. There is minimum loss of materials when nanoparticles with high vapour pressure are produced bv hydrothermal method (Gan et al., 2020).

b)Biosynthesis -

Biological systems like bacteria, fungi, algae and even viruses have been used for synthesis for metallic nanoparticles. The reason this method of synthesis is explored is to reduce the side effects of using chemically synthesised nanoparticles(Thakkar et al., 2009.).Studies have shown that silver nanoparticles are produced intracellularly/extracellularly by bacteria like Pseudomonas stutzeri, Morganella some Lactobacillus strains.* sp. and MKY3(yeast) and fungi like Verticillum, Aspergillus Trichoderma asperellum and

fumigatus are also able to produce silver nanoparticles. Gold nanoparticles are produced when gold ions are supplied to extracts of plants such as *Cymbopogon flexuosus* and *Pelargonium graveolens*.

2.2 Methods for characterization:

X-ray Diffraction -

X-rays have a dual nature(particle/wave) and this is utilized in X-ray diffraction technique. This technique is based on the diffraction of x-rays by nanoparticles and the resulting diffraction pattern is measured by Bragg's law($n\lambda = 2dsin\theta$). It helps in assessing the space between layers of atoms thus determining the crystal structure (Jose Chirayil et al., 2017) In X-ray diffraction technique, the sample is bombarded with a monochromatic beam of X-rays, which get reflected from the sample and are the the detector (Patel detected by and Parsania,2018).

This is an example of X-ray diffraction pattern obtained of ZnO nanoparticles. The peaks obtained determine that it is the hexagonal wurtzite phase of ZnO. No other peaks than that of ZnO indicate that the compound contains no impurities(Talam *et al.*,2012)

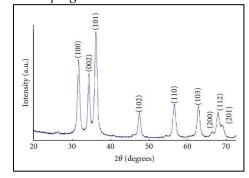


Figure 5: XRD pattern of ZnO nanoparticles

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FOURIER TRANSFORM INFRA-RED SPECTROSCOPY -

In this technique, an infrared spectrum of emission or absorption of the nanoparticles is obtained. A molecule absorbs infrared frequencies equal to its vibrational frequencies. This absorbed energy gets amplified due to the vibrational motion of bonds of the molecule and is given out as radiation to form a spectrum. Fourier transform is the algorithm which converts the data obtained from the interferogram into an actual spectrum. Therefore this technique is used to characterize different elements as well as specific bonds in product nanoparticles(Baudot et al., 2010).

SCANNING ELECTRON MICROSCOPY -

This is a simple method for viewing actual structure as well as measuring the size of the product nanoparticle. In this technique, the sample is bombarded with a primary electron beam. After this beam collides with the sample, secondary electrons and backscatter electrons are created. These electrons are responsible for formation of SEM image(Vladár and Hodoroaba, 2020)

For characterization of the nanoparticle by SEM, the prepared sample should be of good quality; only then will the resulting image will be of high resolution and quality.

SEM image of hydroxy apatite nanoparticles by Kottegoda *et al.* used for characterization of HA nanofertilizer. As can be observed from the figure, the size of the nanoparticles ranges from approximately 17nm to 24nm.

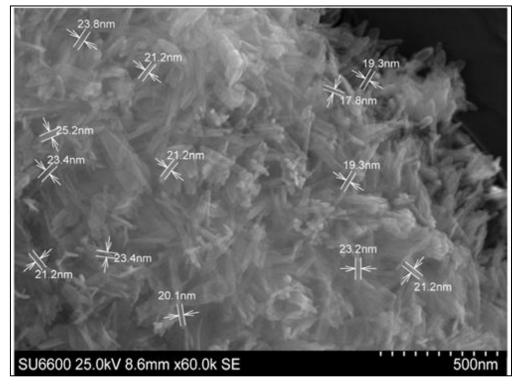


Figure 6: Scanning electron microscope images of synthesized

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- (1) rgonium
- (2) graveolen
- (3) Pelargonium
- (4) graveolen

3. RESULT -

Different nanoparticles used for various crops have been summarized in the

following table. Some of these studies have shown the effect of nanofertilizers when supplied directly in the soil, or by foliar application or in an aqueous solution or tissue culture. Results obtained by hydroponic technique or tissue culture may vary when applied in soil on a larger scale, but they definitely give an idea about the usefulness of nanofertilizers.

Reference	Сгор	Nanofertilizers	Effects	Growth medium
Kottegoda et al.(2011)	No crop, only tested in soil	Urea modified hydroxyapatite nanoparticles encapsu lated under pressure into cavities of the soft wood of <i>Gliricidia sepium</i> .	Slow and sustained release of nitrogen over time	Soil
Shebl <i>et</i> <i>al.</i> (2019)	Cucurbita pepo L	Zinc, Manganese and Iron nanooxides	Improved growth and yield	Soil
Jeyasubra manian <i>et</i> <i>al.</i> (2016)	Spinach	Iron oxide (Fe2O3) nanoparticles	Increased biomass production, stem and root lengths	Aqueous solution
Su <i>et</i> <i>al.</i> (2018)	Peanut	Carbon nanodots	Enhanced stress resistance	Soil
Khodakov skaya et al.(2012)	Tobacco cell culture	Multiwall carbon nanotubes	Increased growth of tobacco cell culture and upregulation of tobacco aquaporin gene	Tissue culture medium
Rangaraj et al.(2012)	Zea mays L.	Porous silica nanoparticles	Increased silica accumulation, leading to regulation of phyto compounds like phenol and proteins.	Soil
Shah & Belezerov a(2009)	Lettuce seeds	Metal (Pd, Cu, Si, Au) nanoparticles	Increased shoot/root ratio	Soil
T. N. V. K. V. Prasad et al.(2012)	Peanut seeds	Zinc oxide(ZnO) nanoparticles	Increased germination, root& shoot growth. Increased pod yield	Soil
Mahajan <i>et al.</i> (2011)	Vigna radiata, Cicer arietinum	ZnO nanoparticles	Increased root and shoot growth at certain concentration	Agar
Salama et al.(2019)	Phaseolus vulgaris	ZnO nanoparticles	Increased concentration of	Soil

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			specific minerals in leaves. Increased amino acid and photosynthetic pigment content	
Rui et al.(2016)	Arachis hypogaea (peanut plant)	Fe2O3 nanoparticles	Increased total Fe content, biomass and chlorophyll content	Soil
Khodakov skaya et al.(2013)	Tomato plant	Carbon nanotubes	Twice the amount of flowers and fruits	Soil
Arora <i>et al.</i> (2012)	Brassica juncea	Gold nanoparticles	Increased growth and seed yield	Soil
Liu and Lal(2014)	Glycine max	Hydroxyapatite nanoparticles	Increased seed yield and biomass production	Soil
Suriyapra bha et al.(2012)	Zea mays L.	Porous silica nanoparticles	Increases concemtration of phytocompounds and silica accumulation in roots	Soil

4. DISCUSSION -

Also the quantity used is less as compared to bulk fertilizers. It is observed from the toxicicity studies that nanoparticles used in proper quantity are not harmful to the environment and hence can easily be used in regular agricultural practices.

Most of these applications are still in the initial stages and have not been used on a large scale. The reason for this may be the relatively higher cost of manufacturing and lesser availability. So it is imperative to find new techniques which will decrease the manufacturing costs, making it more readily available.

5. REFERENCES -

 FAO/WHO expert meeting on the application of nanotechnologies in the food and agriculture sectors: potential food safety implications: meeting report https://www.who.int/publicationsdetail/9789241563932 (accessed Jun 5, 2020).

- (2) Duhan, J. S.; Kumar, R.; Kumar, N.; Kaur, P.; Nehra, K.; Duhan, S. Nanotechnology: The New Perspective in Precision Agriculture. Biotechnol Rep (Amst) 2017, 15, 11–23. https://doi.org/10.1016/j.btre.2017.03.002.
- (3)Iqbal, Muhammad Aamir. (2019). Nano-Fertilizers for Sustainable Crop Production under Changing Climate: A Global Perspective. 10.5772/intechopen.89089.
- (3) Dimkpa, C. O.; Bindraban, P. S. Nanofertilizers: New Products for the Industry? J. Agric. Food Chem. 2018, 66 (26), 6462–6473. https://doi.org/10.1021/acs.jafc.7b02150.
- (4) Wang, X.; Yang, X.; Chen, S.; Li, Q.; Wang, W.; Hou, C.; Gao, X.; Wang, L.; Wang, S. Zinc Oxide Nanoparticles Affect Biomass Accumulation and Photosynthesis in Arabidopsis. Front. Plant Sci. 2016, 6. https://doi.org/10.3389/fpls.2015.01243.
- (5) Rico, C. M.; Majumdar, S.; Duarte-Gardea, M.; Peralta-Videa, J. R.; Gardea-Torresdey, J. L. Interaction of Nanoparticles with Edible Plants

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and Their Possible Implications in the Food Chain I Agric Food Chem 2011, 59 (8), 3485-		14)Baudot, Charles & Tan, Cher & Kong, Jeng. 2010) FTIR spectroscopy as a tool for papo-

nain. J Agric Food Chem 2011, 59 (8), 3485 3498. https://doi.org/10.1021/jf104517j.

- (6) Trakhtenberg, L.; Khatami, N.; Gerasimov, G.; Ilegbusi, O. Effect of Composition and Morphology on Sensor Properties of Aerosol Deposited Nanostructured ZnO+ In2 O3 Films. Materials Sciences and Applications 2015, 6, 220
- (7) Pawar, V.; Kumar, M.; Dubey, P.; Singh, M. K.; Sinha, A.; Singh, P. Influence of Synthesis Route on Structural, Optical, and Electrical Properties of TiO2. Applied Physics A 2019, 125. https://doi.org/10.1007/s00339-019-2948-3.
- (8) Gan, Y. X.; Javatissa, A. H.; Yu, Z.; Chen, X.; Li, M. Hydrothermal Synthesis of Nanomaterials https://www.hindawi.com/journals/jnm/202 0/8917013/ (accessed Jun 4, 2020) https://doi.org/10.1155/2020/89170103.
- (9) Thakkar, K.; Mhatre, S.; Parikh, R. Biological Synthesis of Metallic Nanoparticles. Nanomed Nanotechnol Biol Med 6:257-262. Nanomedicine: nanotechnology, biology, and medicine 2009, 257-262. 6, https://doi.org/10.1016/j.nano.2009.07.002.
- (10) Jose Chiravil, C.; Abraham, J.; Kumar Mishra, R.; George, S. C.; Thomas, S. Instrumental Techniques for the Characterization of Nanoparticles. In Thermal and Rheological Measurement Techniques for Nanomaterials Characterization; Elsevier, 2017; pp 1-36. https://doi.org/10.1016/B978-0-323-46139-9.00001-3.
- (11) Patel, J. P.; Parsania, P. H. 3 Characterization, Testing, and Reinforcing Materials of Biodegradable Composites. In Biodegradable and Biocompatible Polymer Composites; Shimpi, N. G., Ed.; Woodhead Publishing Series in Composites Science and Engineering; pp 55-79. Woodhead Publishing, 2018; https://doi.org/10.1016/B978-0-08-100970-3.00003-1.
- (12) Talam, S.; Karumuri, S. R.; Gunnam, N. Synthesis, Characterization, and Spectroscopic Properties of ZnO Nanoparticles https://www.hindawi.com/journals/isrn/201 2/372505/ (accessed 5, Jun 2020). https://doi.org/10.5402/2012/372505.

es & Tan, Cher & Kong, Jeng. (2010). FTIR spectroscopy as a tool for nanomaterial characterization. Infrared Physics & 434-438. 53. Technology. 10.1016/j.infrared.2010.09.002.

- (15) Vladár, A. E.; Hodoroaba, V.-D. Chapter 3. 2.1.1 - Characterization of Nanoparticles by Scanning Electron Microscopy. In Characterization of Nanoparticles; Hodoroaba, V.-D., Unger, W. E. S., Shard, A. G., Eds.; Micro and Nano Technologies; Elsevier, 2020; pp 7-27. https://doi.org/10.1016/B978-0-12-814182-3.00002-X.
- (16)Kottegoda, Nilwala & Munaweera, Imalka 4. & Adassooriya, Nadeesh & Karunaratne, Veranja. (2011). A green slow-release fertilizer composition based urea-modified on hydroxyapatite nanoparticles encapsulated wood. Current Science. 101. 73-78.
- (17) Shebl, A.; Hassan, A.; Salama, D.; El-Aziz, 5 M. E. A.; Elwahed, M. A. Green Synthesis of Manganese Zinc Ferrite Nanoparticles and Their Application as Nanofertilizers for Cucurbita Pepo L. Beilstein Arch. 2019, 2019 (1), 45. https://doi.org/10.3762/bxiv.2019.45.v1.
- (18) Jeyasubramanian, K.; Thoppey, U. U. G.; 6. Hikku, G. S.; Selvakumar, N.; Subramania, A.; Krishnamoorthy, K. Enhancement in Growth Rate and Productivity of Spinach Grown in Hydroponics with Iron Oxide Nanoparticles. RŠC Adv. 2016, 6 (19), 15451-15459. https://doi.org/10.1039/C5RA23425E.
- 7 (19) Su, L.-X.; Ma, X.-L.; Zhao, K.-K.; Shen, C.-L.; Lou, Q.; Yin, D.-M.; Shan, C.-X. Carbon Nanodots for Enhancing the Stress Resistance of Peanut Plants. ACS Omega 2018, 3 (12), 17770-17777.

https://doi.org/10.1021/acsomega.8b02604.

- (20) Khodakovskaya, M. V.; de Silva, K.; Biris, 8. A. S.; Dervishi, E.; Villagarcia, H. Carbon Nanotubes Induce Growth Enhancement of Tobacco Cells. ACS Nano 2012, 6 (3), 2128-2135. https://doi.org/10.1021/nn204643g.
- 9. (21) Rangaraj, S.; Gopalu, K.; R, Y.; Periasamy, P.; VENKATACHALAM, R.; Kannan, N. Growth and Physiological Responses of Maize (Zea Mays L.) to Porous Silica Nanoparticles in

Research & Reviews in Biotechnology & Biosciences Website: www.biotechjournal.in Volume-8, Issue No: 1, Year: 2021 (January-June) DOI: http://doi.org/10.5281/zenodo.5118413

Soil. Journal of Nanoparticle Research 2012, 14. https://doi.org/10.1007/s11051-012-1294-6.

- 10. (22) Shah, V.; Belozerova, I. Influence of Metal Nanoparticles on the Soil Microbial Community and Germination of Lettuce Seeds. 2009.
- 11. (23) T. N. V. K. V. Prasad, P. Sudhakar, Y. Sreenivasulu, P. Latha, V. Munaswamy, K. Raja Reddy, T. S. Sreeprasad, P. R. Sajanlal & T. Pradeep (2012) EFFECT OF NANOSCALE ZINC OXIDE PARTICLES ON THE GERMINATION, GROWTH AND YIELD OF PEANUT, Journal of Plant Nutrition, 35:6, 905-927, DOI: 10.1080/01904167.2012.663443
- (24) Mahajan, P.; Dhoke, S. K.; Khanna, A. S. Effect of Nano-ZnO Particle Suspension on Growth of Mung (Vigna radiata) and Gram (Cicer arietinum) Seedlings Using Plant Agar Method https://www.hindawi.com/journals/jnt/2011

/696535/ (accessed Jun 3, 2020). https://doi.org/10.1155/2011/696535.

- (25) Salama, D. M.; Osman, S. A.; Abd El-Aziz, M. E.; Abd Elwahed, M. S. A.; Shaaban, E. A. Effect of Zinc Oxide Nanoparticles on the Growth, Genomic DNA, Production and the Quality of Common Dry Bean (Phaseolus Vulgaris). Biocatalysis and Agricultural Biotechnology 2019, 18, 101083. https://doi.org/10.1016/j.bcab.2019.101083.
- (26) Rui, M.; Ma, C.; Hao, Y.; Guo, J.; Rui, Y.; Tang, X.; Zhao, Q.; Fan, X.; Zhang, Z.; Hou, T.; Zhu, S. Iron Oxide Nanoparticles as a Potential Iron Fertilizer for Peanut (Arachis Hypogaea). Front. Plant Sci. 2016, 7. https://doi.org/10.3389/fpls.2016.00815.
- (27) Khodakovskaya, M. V.; Kim, B.-S.; Kim, J. N.; Alimohammadi, M.; Dervishi, E.; Mustafa, T.; Cernigla, C. E. Carbon Nanotubes as Plant Growth Regulators: Effects on Tomato Growth, Reproductive System, and Soil Microbial Community. Small 2013, 9 (1), 115–123. https://doi.org/10.1002/smll.201201225.
- 16. (28)Arora, S., Sharma, P., Kumar, S. et al. Goldnanoparticle induced enhancement in growth and seed yield of Brassica juncea . Plant Growth Regul 66, 303–310 (2012). https://doi.org/10.1007/s10725-011-9649-z

- (29) Liu, R.; Lal, R. Synthetic Apatite Nanoparticles as a Phosphorus Fertilizer for Soybean (Glycine Max). Sci Rep 2014, 4. https://doi.org/10.1038/srep05686.
- (30) Suriyaprabha, R., Karunakaran, G., Yuvakkumar, R. et al. Growth and physiological responses of maize (Zea mays L.) to porous silica nanoparticles in soil. J Nanopart Res 14, 1294 (2012). https://doi.org/10.1007/s11051-012-1294-6